

User Guide to Services

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0. Introduction

Severn Pathology, North Bristol NHS Trust provides a Cellular Pathology service from a laboratory based in Phase two, Pathology Sciences, Southmead Hospital. There is an essential services laboratory (ESL) located in UHBristol.

The Department provides a wide-ranging and comprehensive diagnostic Histopathology, diagnostic Cytopathology and Cervical screening service.

Services offered include consultation with clinical colleagues to assist in the interpretation of reports and provide advice about the collection, handling, fixation and submissions of specimens for investigation.

The department manages two mortuary facilities at Southmead Hospital and in St Michael's Hospital. Since April 2009, when the Bristol City Council / Coroner's mortuary opened at Flax Bourton, the service in Southmead is limited to storage and management of the deceased and the facilitation of viewings. A paediatric/perinatal mortuary service is provided at St Michael's Hospital. **The department has no responsibility for the mortuary on Level 4, Queen's Building, BRI or the body store in the Bristol Children's Hospital.**

In April 2019 the department was awarded the contract for the provision of Laboratory Services to support the delivery of the HPV Primary Screening Pathway within the NHSCSP to the population of the South West of England. There will be a phased transfer of services from the cytology laboratories across the South West region, resulting in NBT processing all cervical samples taken in the South West of England by March 2020. NBT also provides a cervical screening service to Spire Healthcare, the Isle of Man and the Bailiwick of Guernsey. There is currently an annual workload of ~120,000 cervical screening samples and from 1st April 2020 this is expected to rise to ~300,000. All service users have fully converted to the HPV Primary screening pathway and samples are processed using Hologic Aptima HPV assay, followed by the screening of high risk HPV detected samples using Hologic Thinprep slide preparation technology. Further details on service provision, supplies and arrangements during the transitional phase can be obtained by contacting the cytopathology section.

1. Working Hours

The Department is open to receive specimens and offer technical advice Monday – Friday, 07:00 – 18:00. The Histology laboratory also has a Saturday service which runs from 09:00 – 17:00.

Consultant Pathologists are available for advice and opinion Monday to Friday 09:00 – 17:00. Outside of these hours, contact with the renal on-call Consultant can be made via switch board. **Please note there is no general Consultant on-call.**

2. Specimen Identification

With the exception of intraoperative frozen section requests, requests for Histopathology or Cytopathology can be made on ICE, this is the preferred and safest method of requesting pathology. Where users are unable to access ICE, it is essential that a completed request form accompany the specimen. Specimen containers (not the lid) must be appropriately labelled in accordance with NBT Clinical Governance Policy CG45: **Policy for Request Form and Specimen Labelling.**

Specimens **MUST** bear,

- Patient number (e.g. NHS number, hospital number, ICE number)
- Date of birth
- Family name and first name
- Description of samples

Failure to provide this information may delay or prevent the processing of a specimen

In addition to the above, Histology Request forms should bear the following information,

- Patient address with postcode
- Requestor and Consultant name and location
- Date and time specimen was taken
- Biopsy site. This is particularly important if multiple samples are to be sent.
- Relevant clinical details including LMP as appropriate, details of any previous biopsies
- Patient category (NHS / Private / Other)

Request forms for cervical screening should be the printed pre-populated Open Exeter A4 portrait editable HMR5 form. Please see FAQs on website.

Cervical screening forms also need a registered sample taker code. Registration is available at: <http://extra.scwcsu.nhs.uk/screeningimmunisation/Cervical/main2.htm>

Failure to provide the information may result in delays in producing a report.

3. Health and Safety

Specimens

(a) Any patient with or suspected any of the following are considered to be an Inoculation Risk,

- HIV
- Hepatitis B
- Hepatitis C
- Creutzfeld Jakob
- Tuberculosis

(b) For the safety of staff, please ensure that investigations are kept to a minimum.

(c) Forms must be clearly labelled with a “**Biohazard Risk**” or “**Inoculation Risk**” sticker with, **the nature of the risk given on the request form.**

(d) If a patient having an Inoculation Risk is admitted to a trust hospital, the Consultant Microbiologist should be advised as soon as possible.

(e) Specimen transport from the Brunel building to Cellular Pathology is via the porters.

(f) In Southmead Hospital, to control and minimise exposure to histology fixative reagents the TissueSafe system is available for use for Theatre samples from the Brunel building.

(g) Safe transport of specimens from the Brunel Building to Cellular Pathology:-

Cellular Pathology specimens must NOT be placed in the 'POD' transport system

Formalin

Formalin is a hazardous substance and should only be handled wearing appropriate PPE (gloves, eye protection, apron/ scrubs).

Pots should be stored at room temperature in a safe and upright position to minimise the risk of spillage/ leakage.

Should a spillage occur:

- For small volumes (up to approx. 100ml) – ventilate the area where possible and use absorbant material (i.e. paper towel) to clear up spillage.
- For larger volumes (>100ml) – Use a spill kit where available following the provided instructions. If no spill kit is available restrict access to the area and call the laboratory immediately for advice (0117 4149800).

LBC Pots

Contents are flammable so should be stored at room temperature (15-30C) and away from potential ignition sources.

Spillages can be dealt with using absorbant material i.e. paper towel.

4. Transport

- (a) In Southmead hospital, specimen transport to Cellular Pathology is via the porters. Please check with your location for details on how this is arranged or by contacting Facilities and Estates.
- (b) Samples from UHBristol sites are delivered by porter to Pathology reception on level 8 of the BRI. These samples are then transported to Southmead via regular City Sprint transports.
- (c) Samples from Weston-Super-Mare are delivered to Pathology at WGH and are transported to Southmead via City Sprint transports.
- (d) City Sprint under contract to Pathology Sciences NBT provides a routine service to local hospitals and GP surgeries.
- (e) Cervical Cytology from outside of the NBT transport catchment will be collected by transports operated by UHBristol or Weston Area Health or by City Sprint.
- (f) Transport of urgent frozen section samples from UHB to Southmead hospital is via ambulance from the BRI. Details can be found on the UHB intranet site or from theatre managers. Contact the department for further details.
- (g) For emergency transport, contact the Trust Transport Department on 0117 414 5423.

5. Supplies

Supplies for use in collection and despatch of Histology and non-Gynae cytology samples can be ordered through Pathology Sciences.

Southmead E-mail:

PathologyConsumablesSouthmead@nbt.nhs.uk

Requests can be made using a pro forma list and supplies are dispatched by the Trust Transport system within 3 days.

Supplies of larger (1L+) pots both with and without formalin are available directly from the Histology laboratory at Southmead hospital or from the ESL laboratory for UHB sites. Please contact the department if you require further information. All formalin supplies should be stored at room temperature in a safe upright position where the possibility of spillage is minimised.

LBC supplies are available from an external company 'CellPath' please contact info@cellpath.co.uk or call 01686 611333.

- If you have stock that has expired please contact the appropriate laboratory for disposal advice.
- ALWAYS CHECK YOUR SUPPLIES ARE IN DATE BEFORE USE. Expired reagents may affect results.

6. Key Contacts and Consultant staff

For guidance and advice about any aspects of the work of the department, please contact an appropriate member of staff.

SMD: Pathology Sciences

Name	Tel	Fax
Dr Nidhi Bhatt Consultant Cellular Pathologist Head of Department	0117 41-49880 Secretary: - 0117 41-49890	
Mr Andrew Heryet Laboratory Manager	0117 41-49849	
Mr Mark Orrell Operations Manager	0117 41-49875	

Histopathology - Enquiries & Results	0117 41-49890	0117 41-49390
Cytopathology - General Enquiries	0117 41-49889	0117 41-49389

Dr Zainab Abdul-Rahman Consultant Cellular Pathologist	0117 41-49832	
Dr Elizabeth Aliaga Consultant Cellular Pathologist	0117 41-49862	
Dr Kavita Amarasinghe Consultant Cellular Pathologist	0117 41-49829	
Dr Nahida Banu Consultant Cellular Pathologist	0117 41-49837	
Dr Andrew Bamber Consultant Cellular Pathologist	0117 41-49827	
Dr Sophie Beavers Consultant Cellular Pathologist	0117 41-49869	
Dr Naomi Carson Consultant Cellular Pathologist	0117 41-49868	
Dr Anastasios Chatzitoliou Consultant Cellular Pathologist	0117 41-49824	
Dr Richard Daly Consultant Cellular Pathologist	0117 41-49878	
Dr Karin Denton Consultant Cellular Pathologist	0117 41-49828	
Dr Mohammed Khan Consultant Cellular Pathologist	0117 41-49835	
Dr Francesca Maggiani Consultant Cellular Pathologist	0117 41-49877	
Dr Zsombor Melegh Consultant Cellular Pathologist	0117 41-49884	
Dr Behrang Mozayani Consultant Cellular Pathologist	0117 41-49825	
Dr Timothy Murigu Consultant Cellular Pathologist	0117 41-49866	
Dr Jon Oxley Consultant Cellular Pathologist	0117 41-49831	
Dr Joya Pawade Consultant Cellular Pathologist	0117 41-49886	
Dr Demetris Poyiatzis Consultant Cellular Pathologist	0117 41-49840	
Professor Colin Purdie Consultant Cellular Pathologist	0117 41-49908	
Dr Mohammed Sohail Consultant Cellular Pathologist	0117 41-49836	
Dr Judit Sutak Consultant Cellular Pathologist	0117 41-49838	
Dr Eleni Toumazou Consultant Cellular Pathologist	0117 41-49876	
Dr Newton Wong Consultant Cellular Pathologist	0117 41-49883	
Dr Yve Zhang Consultant Cellular Pathologist	0117 41-49885	

Helen Burrell Consultant BMS	0117 41-49806	
Nichole Villeneuve Consultant BMS	0117 41-49889	
Lynne Williams Consultant BMS	0117-41-49969	

BRI: Essential Services Laboratory

Name	Tel	Fax
Dr Miranda Pring Consultant Cellular Pathologist	0117 34-29500	
Dr Ed Sheffield Consultant Cellular Pathologist	0117 34-27622	
Laboratory Enquiries	0117 34-27556	
Report Enquiries	0117 41-49890	

7. GP Access

- (a) GP enquiries are welcomed and can be made by telephoning during normal working hours (09.00 – 17.00 hrs).
- (b) Reports are dispatched by hospital postal service (the majority GP practices have direct access to results via computer).

8. Histopathology specimens

- (a) Routine specimens – where fresh tissue is not required

- Should be placed in at least 10 times their volume of fixative (neutral buffered formalin) in an appropriate container. Specimen containers and fixative will be supplied on request. All samples are to be accompanied by a fully completed Request Form or ICE request.
- Delayed fixation, inadequate fixation (through insufficient volume of fixative), and sample handling (i.e. crush artefact, diathermy etc.) may have a detrimental affect on the initial diagnosis and may invalidate subsequent tests such as immunohistochemistry.
- Formalin is a hazardous substance and care should be taken when handling (see H&S above). The lids of all specimen containers must be securely applied to prevent leakage.
- All samples should be transported within specimen transport boxes (i.e. Daniel's boxes or equivalent) wherever possible. Those that are too large to fit into a transport box must be double bagged.

- Small specimen pots should be placed into the sealed plastic specimen bags with the request form (if applicable) placed in the front envelope of the bag. This is to ensure the request forms are kept clean and avoid being ruined by potential formalin leaks.
- Large, operative specimens collected during working hours should be sent in a sealed box, or specimen bucket, unfixated to the laboratory with a porter to be received by 17:00. After 17:00, unless prior specific arrangements have been made, place specimen into fixative and transport to the laboratory as usual.
- Specimens packaged using the TissueSafe system must be transported in the appropriate coolbox with temperature logger. If these samples cannot be delivered to the laboratory by 17:00 they should be refrigerated overnight and delivered as soon as possible the following day.
- All Inoculation Risk specimens must be fixed in formalin and labelled appropriately.
- Urgent specimens may be able to be processed (if of a suitable size) within the day it is taken. Specimens must be in the department before 12.30 pm. Please contact the department to discuss.
- For samples where rapid diagnosis is required for clinical reasons, these should be placed into formalin and promptly sent to the laboratory, marked URGENT, and accompanied by a request form bearing appropriate details.

(b) It is the intention of the department that most routine biopsies to be available on the ward computer within seven working days of receipt to the laboratory. This does however depend upon the exact nature of the specimen and the requirement for additional processing, testing or opinion. Times may also be extended due to availability of staff.

(c) If immunofluorescence studies are required, Michel's transport medium can be supplied. Please telephone the department to discuss.

(d) All histology is reported or personally supervised by consultants.

(e) Frozen Sections

- Samples for frozen section diagnosis at Southmead Hospital will be accepted at any time during working hours, 9.00 am to 4.30 pm. Service outside these hours is by prior arrangement only. **24 hours notice is required to ensure appropriate staff members are available.** Please telephone the department with details ext. 49890. For requests originating from areas within UHBristol, please phone the ESL on ext. 27556.

[9. Diagnostic Cytopathology specimens](#)

(a) All samples, specimen pots or slides must be clearly labelled and include the patient name, hospital number and date of birth including any treatment that the patient is receiving. These must be accompanied by a request form or ICE request which should include relevant medical details, including the sample site. Requests for breast aspiration cytology should state if the patient is pregnant or on contraceptives.

(b) All specimens that may be an inoculation risk should be labelled as such (Please see "(3) Health and Safety" above).

(c) The department aims to be able to issue at least a preliminary report on all cases within 5 days. This does however depend upon the exact nature of the specimen and the requirement for additional processing, testing or opinion. If a rapid result is required specimens should be promptly sent to the laboratory, marked URGENT, and accompanied by a request form bearing appropriate details.

(d) One-stop clinics in support of the Head & Neck service are held weekly at the Bristol Dental Hospital and are attended by a Consultant Pathologist and BMS in order to facilitate rapid diagnosis.

(e) Please note that Cerebrospinal fluids (CSF) samples are handled by Neuropathology (see details below).

(f) Specimen Collection

i) Cellular Pathology

Specimen type	Requirements	Quality Measures
Serous Fluids (pleural, ascitic, pericardial, peritoneal lavage/washings)	Serous fluids are collected into plain universals.	
Cyst Fluids	Collect into dry, sterile universals.	
Bronchoalveolar Lavage (BAL) / Bronchial Washings	Collect into dry, sterile universals. UHB Paediatric BALs: Place sample in a clear plastic specimen bag labelled 'urgent'. Send sample via porter to the Cellular Pathology laboratory (ESL) on level 8 of the BRI. For samples taken on a Friday please ring the Cytology laboratory on 0117 414 9889 to alert us to the arrival.	
Fine Needle Aspiration (FNA) & Needle Washings	Using pencil , label up to five clean, frosted end microscope slides* with: (a) Patient's surname (b) Patient's forename and (c) Patient's DOB or hospital number. Place 1 drop of specimen near bottom of each slide and spread gently using another clean slide. Rapidly air-dry the slides, prior to placing in a plastic slide container. NB; If squamous cell carcinoma is suspected, at least one slide should be spray fixed and labelled as such. Flush needle into a bottle containing 5ml Cytolyt and send the needle washings together with the FNA slides (do not send the needle).	If the sample is too thick, it may not be possible to obtain a diagnosis. Delay in air drying or fixation affects cellular preservation and can affect diagnosis. Fixed slides must immediately be fixed with Cytospray* cytology fixative to prevent air-drying.

Breast Fine Needle Aspiration (FNA)	Breast FNAs should be spread thinly onto a clean labelled slide (see FNA above) and air-dried rapidly. Any needle washings should be placed in a sterile universal containing 5ml Cytolyt (available from Cytology x5643)	If the sample is too thick, it may not be possible to obtain a diagnosis. Delay in air drying or fixation affects cellular preservation and can affect diagnosis.
Brushings (bronchial, oesophageal, biliary, gastric)	A minimum of two slides are required. Using pencil , label clean, frosted end microscope slides* with: (a) Patient's surname (b) Patient's forename and (c) Patient's DOB or hospital number. Transfer material from brush onto slides immediately after collection, using a gentle but firm rolling technique. One slide is rapidly air-dried and the other spray fixed. Slides should be allowed to dry, labelled with identification 'fix' or 'dry' and placed in a plastic slide container. We also receive brush tips in 20ml Cytolyt.	Delay in air drying or fixation affects cellular preservation and can affect diagnosis. Fixed slides must immediately be fixed with Cytospray* cytology fixative to prevent air-drying.
Synovial Fluid	Collect into dry, sterile universals. Samples must be sent to Cytology department as soon as possible .	Incorrect storage can lead to the formation of contaminating crystals.
Urine / Urinary Tract Washings	Place in bottles containing alcohol.* Urine sample should ideally be an aliquot of the whole output, although a midstream sample is acceptable. An early morning sample is not appropriate. Please indicate on the request form if specimen is a catheter or cystoscopy sample.	No alcohol can lead to the degeneration of the cellular content and to bacterial overgrowth and can affect diagnosis.
Sputum	Collect into dry, sterile sputum pots. The specimen should be collected before any food is eaten and before teeth are cleaned and should be a deep cough, early morning specimen taken with the assistance of a physiotherapist if needed. Saliva is not adequate.	

* Available by contacting Cytology, Southmead Hospital on ext. 49889

ii) Neuropathology

Specimen type	Requirements	
Cerebrospinal fluid (CSF)	Contact Neuropathology who provide this service in support of Neurosciences	0117 41-42400

All samples need to be sent to the laboratory **as soon as possible** and arrive at the laboratory during routine opening hours. If this is not possible store in the fridge and send in on the next working day. Any delay can lead to the cellular content degenerating.

10. [Cytopathology - Cervical Screening](#)

All samples for cervical screening must be collected into ThinPrep vials and be sent to the laboratory in the orange bags on the next available transport.

All requests for screening will need to be made via the Open Exeter IT system by filling and printing the Open Exeter HMR101 form and including it with the sample. This will ensure correct patient identification, correct patient management and improve efficiency for the whole service.

- All samples for cervical screening must be collected into ThinPrep vials and be sent to the laboratory with a typed HMR101 request form.
- Cervical screening results are available within 14 days of the sample being taken.

11. [SMD Brunel Building Mortuary](#)

The Mortuary is staffed Monday to Friday, 08.00 to 16.00 hours.
 A member of staff is available on standby at all other times.

(a) Requests for viewings of deceased by relatives and friends should be directed to the Mortuary: 0117-414 1700, or 0117-414 1701.
 Requests for viewings during stand-by periods should be directed to the on-call technician via Trust Switchboard.

(b) Note that no Post Mortem examinations are carried out on NBT premises. All adult Coroner's PMs will be transferred to the facility at Flax Bourton. North Bristol Trust no longer provides Adult Hospital Post *Mortem Examinations*. *Perinatal* and Neonatal deaths that are consented for PM will be transferred to the Mortuary at St Michael's Hospital. Neonatal deaths that have a Coroner's Post Mortem will be transferred to a facility determined by the Coroner's Office.

(c) Reporting deaths to Coroner

For further information contact the bereavement office: 0117-414 0180, 414 0182 or 414 0183

(d) General enquiries

Contact the Mortuary.

12. St Michaels Hospital Paediatric Mortuary

The Paediatric Mortuary located at St Michaels Hospital can be contacted on 0117 34-25428. It is open Monday to Friday 09.00 to 16.30 hours. Stand-by service is provided by the UH Bristol Mortuary Team.

(a) Perinatal and Neonatal Hospital Post Mortems

Written consent must be given for a post mortem examination. Clinicians and Specialist Midwives consenting for Perinatal and Neonatal PMs must have undertaken PM Consent Training / update within the last 2 years.

(b) Coroner's Post Mortems. Neonatal deaths that have a Coroner's Post Mortem will be transferred to a facility determined by the Coroner's Office.

13. Referral Services

(a) The department is able to consider offer technical services to other Cellular Pathology departments. Please contact the Laboratory Manager or Operations Manager to discuss requirements.

Potential users should be aware that the department cannot validate tissue fixation and processing of externally referred tissue blocks.

Techniques used are appropriately verified and subject to appropriate QA and QC.

Please note that the department uses a small number of research use only (RUO) antibodies. These antibodies have been verified in house, to reflect clinical use. However the expected staining characteristics have not been formally validated by the manufacturer. These antibodies are never used in isolation and are not the sole means of diagnosis for the cases they are requested on. We are committed to using CE-IVD approved methodologies whenever possible and update our antibody stock when these become available

(b) Details are provided below of the accredited referral laboratories and services used by the department.

Name and Address of Department	Named Consultant (if applicable)	Repertoire / Scope of Service to be provided
Source Bioscience, 1 Orchard Place, Nottingham, NG8 6PX	-	Reporting of Histology specimens including Breast pathology
LD Path	-	Reporting of Histology specimens
Health Services Laboratories Advanced Diagnostics, The Halo Building, 1 Mabledon Place, London, WC1H 9AX	-	Immunohistochemistry (requests which fall outside the current repertoire)

UCL, 21 University St. London WC1E 6JJ	Professor M Novelli	Second Opinion
UCL, 21 University St. London WC1E 6JJ	Dr A Ramsay	Second Opinion
Dept of Medicine, School of Medicine, Cardiff University, Heath Park, Cardiff, CF14 4XN	-	Specialist IHC
University Hospital of Wales, Cardiff	-	Electron Microscopy: preparation of digital images for reporting
Oxford Radcliffe Hospital	Dr Ian Roberts	Reporting Renal biopsies
Oxford Radcliffe Hospital	Dr F Pezzello	Reporting / Second Opinion, Haem malignancies
Frimley Park, Camberley, Surrey, GU16 7UJ	Professor Cook	Dermatopathology
Gloucestershire Hospitals	Prof N Shepherd	Second Opinion, GI specimens
Royal Orthopaedic Hospital, Birmingham	Professor Kindblom / Dr Sumathi	Second Opinion, Sarcoma
Royal Marsden, Dows Road, Sutton, Surrey	Professor Martin Gore Professor C Fisher	Second Opinion
Guy's and St Thomas, London (Viapath)	-	Gene Analysis
Guy's and St Thomas, London (Viapath)	Dr Bridget Wilkins	Second Opinion, Lymphoma
Guy's and St Thomas, London (Viapath)	Dr Eduardo Calonje	Second Opinion, Dermatopathology
Institute of Liver Studies, Kings College Hospital	Dr Alex Knisely and colleagues	Liver biopsies
University Hospital Birmingham	Prof Stefan Hubscher	Liver biopsies
Newcastle on Tyne NHS FT	Dr Sarah Johnson	Second Opinion
Royal National Orthopaedic Centre, Stanmore	Dr Fernanda Amary	Second Opinion
Royal Free Hospital	Professor A P Dhillon	Second Opinion
Christie Hospital, Manchester	Dr Shanks	Second Opinion
Edinburgh RI (NHS Lothian University Hospitals)	Dr William Wallace	Second Opinion
University Hospital of Wales, Cardiff	Dr Stefan Dojcinov Dr Kenneth May	Second Opinion
Nuffield Orthopaedic Centre, Oxford (Part of Oxford University Hospitals NHS Trust)	Professor Athanasou	Second Opinion, Bone and soft tissue tumours
Oxford Radcliffe Hospital	Dr Lisa Browning	Urology
Cardiff	Dr Murali Varma	Urology

Royal Brompton	Prof Andrew Nicholson	Expert opinion, respiratory
Edinburgh	Dr Chris Bellamy	Liver
Liverpool	Professor Fiona Campbell	Pancreaticobiliary pathology
Liverpool	Professor Sarah Coupland	Reporting ocular specimens
University Hospital of Wales, Cardiff	Dr Sergey Popov	Second opinion paediatric pathology
Birmingham Children's Hospital	Dr Rachel Brown	Paediatric Liver Pathology
University Hospitals Birmingham	Dr Philippe Taniere	ROS1 testing
Great Ormond Street Hospital	Dr Michael Ashworth	Cardiac Pathology
UMC Utrecht, Netherlands	Dr Peter Nikkels	Fetal Pathology/Skeletal dysplasias
Manchester Children's Hospital	Dr Anna Kelsey	Paediatric Surgical Pathology
Manchester Children's Hospital	Dr Ed Cheesman	Paediatric Surgical Pathology
Wales Institute in Forensic Medicine	Dr Steve Leadbeatter	Paediatric Forensic Pathology
St James University Hospital Leeds	Dr Andrew Hanby	Breast second opinion
Belfast	Prof Glenn McCluggage	Second opinion, Gynae

This does not represent an exhaustive list and other laboratories or named consultants may be referred to in order to obtain tests and opinions that best meet the needs of users and patients

[14. Complaints](#)

In the event that users have to cause to raise a complaint about the service provided, please contact the Clinical Head of Department or Laboratory Manager in first instance, contact details in section 6 of this document. Alternatively, a complaint can be raised using the Trust's procedure available [here](#).

[15. Protection of personal information](#)

The Department of Cellular Pathology operates within the Information Governance policies and procedures laid down by North Bristol NHS Trust. Further details are available [here](#).

[16. Consent](#)

Several hundred Histopathology and Cytology requests are received in the department each day. For some rare or complex tests patient, specimens may be sent to specialist laboratories elsewhere which have the necessary expertise or an expanded repertoire of tests. In some cases there will be only one specialist laboratory in the whole country which performs a particular test, meaning using referral laboratories is essential.

The department has a policy and procedure in place to govern how we choose these referral laboratories. They are selected for their expertise and their quality standards and regular checks are made on their accreditation status, which gives us assurance that they have procedures in place for

the protection of information.

When specimens are sent to a referral laboratory we need to send some 'patient identifiers' such as name and date of birth. In some tests it is essential to send further clinical information to allow the referral laboratory to interpret the results for the individual patient.

Consent to a specimen being taken and analysed is implied by the patient presenting to the point of specimen collection. The responsibility for obtaining informed consent for the test(s) resides with the individual ordering the test. Informed consent should cover all the tests being done, implications of their results and disclosure of clinical and personal details to personnel (in the requesting organisation and any other healthcare organisations involved in providing the test).